What is claimed is:

1. A method of removing small negatively charged organic molecules from a biological sample mixture, the method comprising:

providing a surface comprising an anion exchange material;

providing a biological sample mixture comprising small negatively charged organic molecules having a molecular weight of less than about 6,000; wherein the biological sample mixture is selected from the group consisting of a nucleic acid amplification reaction mixture and a nucleic acid labeling reaction mixture; and

contacting the biological sample mixture with the surface comprising the anion exchange material to remove at least a portion of the small negatively charged organic molecules from the biological sample mixture.

2. A method of removing small negatively charged organic molecules from a biological sample mixture, the method comprising:

providing a surface comprising an anion exchange material partially coated with a negatively charged polymer;

providing a biological sample mixture; and

contacting the biological sample mixture with the surface comprising an anion exchange material partially coated with a negatively charged polymer to remove at least a portion of the small negatively charged organic molecules from the biological sample mixture.

3. The method of claim 2 wherein the negatively charged polymer is a polyelectrolyte.

4. The method of claim 3 wherein the negatively charged polyelectrolyte is selected from the group consisting of a polystryene sulfonic acid, polyvinyl phosphonic acid, polyvinyl boric acid, polyvinyl sulfonic acid, polyvinyl sulfuric acid, polystyrene phosphonic acid, polyacrylic acid, polymethacrylic acid, lignosulfonate, carrageenan, heparin, chondritin sulfate, salts thereof, and mixtures thereof.

5. The method of claim 2 wherein the anion exchange material comprises quaternized nitrogen.

6. The method of claim 2 wherein the biological sample mixture is a nucleic acid sequencing reaction mixture.

7. The method of claim 6 wherein the small negatively charged organic molecules are selected from the group consisting of dye-labeled terminators, primers, degraded dye molecules, deoxynucleotide triphosphates, and mixtures thereof.

8. The method of claim 7 wherein the small negatively charged organic molecules comprise dye-labeled terminators.

9. The method of claim 8 wherein the dye-labeled terminators are selected from the group consisting of dideoxynucleotide triphosphates, dideoxynucleotide diphosphates, dideoxynucleotide monophosphates, dideoxynucleosides, and combinations thereof.

10. The method of claim 8 wherein contacting the biological sample mixture with the surface comprising an anion exchange material partially coated with a negatively charged polymer is carried out under conditions effective to remove substantially all the dye-labeled terminators from the biological sample mixture.

11. The method of claim 2 wherein the biological sample mixture is a PCR reaction mixture.

12. The method of claim 11 wherein the small negatively charged organic molecules are selected from the group consisting of primers, degraded dye molecules, deoxynucleotide triphosphates, and mixtures thereof.

13. The method of claim 12 wherein contacting the biological sample mixture with the surface comprising an anion exchange material partially coated with a negatively charged polymer is carried out under conditions effective to remove substantially all the primers from the biological sample mixture.

14. The method of claim 2 wherein the small negatively charged organic molecules have a molecular weight of less than about 6,000.

15. The method of claim 2 wherein contacting the biological sample mixture with the surface comprising an anion exchange material partially coated with a negatively charged polymer comprises agitating while contacting.

16. The method of claim 2 which is carried out in a microfluidic device.

17. A method of removing small negatively charged organic molecules from a biological sample mixture, the method comprising:

providing a surface comprising quaternary ammonium ions partially coated with a negatively charged polyelectrolyte;

providing a biological sample mixture; and

contacting the biological sample mixture with the surface comprising quaternary ammonium ions partially coated with a negatively charged polyelectrolyte to remove at least a portion of the small negatively charged organic molecules from the biological sample mixture.

18. A method of removing small negatively charged organic molecules from a biological sample mixture, the method comprising:

providing a surface comprising quaternary ammonium ions partially coated with a negatively charged polyelectrolyte;

providing a biological sample mixture; and

contacting the biological sample mixture with the surface comprising quaternary ammonium ions partially coated with a negatively charged

polyelectrolyte to remove at least a portion of the small negatively charged organic molecules from the biological sample mixture;

wherein the biological sample mixture comprises a nucleic acid amplification reaction mixture.

19. The method of claim 18 which is carried out in a microfluidic device.

20. A method of removing small negatively charged organic molecules from a biological sample mixture, the method comprising:

providing a device comprising at least one process array that comprises a surface comprising an anion exchange material;

providing a biological sample mixture in the at least one process array, wherein the biological sample mixture comprises small negatively charged organic molecules having a molecular weight of less than about 6,000; and

transferring the biological sample mixture within the at least one process array, wherein the biological sample mixture and the surface comprising an anion exchange material remain in contact for a sufficient time to remove at least a portion of the small negatively charged organic molecules from the biological sample mixture.

21. A method of removing small negatively charged organic molecules from a biological sample mixture, the method comprising:

providing a device comprising at least one process array that comprises a surface comprising an anion exchange material partially coated with a negatively charged polymer;

providing a biological sample mixture in the at least one process array; and

transferring the biological sample mixture within the at least one process array, wherein the biological sample mixture and the surface comprising an anion exchange material partially coated with a negatively charged polymer remain in contact for a sufficient time to remove at least a portion of the small negatively charged organic molecules from the biological sample mixture.

22. The method of claim 21 wherein the negatively charged polymer is a polyelectrolyte.

23. The method of claim 22 wherein the negatively charged polyelectrolyte is selected from the group consisting of a polystryene sulfonic acid, polyvinyl phosphonic acid, polyvinyl boric acid, polyvinyl sulfonic acid, polyvinyl sulfuric acid, polystyrene phosphonic acid, polyacrylic acid, polymethacrylic acid, lignosulfonate, carrageenan, heparin, chondritin sulfate, salts thereof, and mixtures thereof.

24. The method of claim 21 wherein the anion exchange material comprises quaternary ammonium ions.

25. The method of claim 21 wherein the biological sample mixture is a nucleic acid sequencing reaction mixture.

26. The method of claim 25 wherein the small negatively charged organic molecules are selected from the group consisting of dye-labeled terminators, primers, degraded dye molecules, deoxynucleotide triphosphates, and mixtures thereof.

27. The method of claim 26 wherein the small negatively charged organic molecules comprise dye-labeled terminators.

28. The method of claim 27 wherein the dye-labeled terminators are selected from the group consisting of dideoxynucleotide triphosphates, dideoxynucleotide diphosphates, dideoxynucleotide monophosphates, dideoxynucleosides, and combinations thereof.

29. The method of claim 27 wherein the biological sample mixture and the surface comprising an anion exchange material partially coated with a

negatively charged polymer are contacted under conditions effective to remove substantially all the dye-labeled terminators from the biological sample mixture.

30. The method of claim 21 wherein the biological sample mixture is a PCR reaction mixture.

31. The method of claim 30 wherein the small negatively charged organic molecules are selected from the group consisting of primers, degraded dye molecules, deoxynucleotide triphosphates, and mixtures thereof.

32. The method of claim 31 wherein the biological sample mixture and the surface comprising an anion exchange material partially coated with a negatively charged polymer are contacted under conditions effective to remove substantially all the primers from the biological sample mixture.

33. The method of claim 21 wherein the small negatively charged organic molecules have a molecular weight of less than about 6,000.

34. The method of claim 21 wherein the biological sample mixture and the surface comprising an anion exchange material partially coated with a negatively charged polymer are agitated while in contact.

35. The method of claim 21 wherein the at least one process array comprises a loading chamber, at least one process chamber, and at least one distribution channel connecting the loading chamber and the at least one process chamber.

36. A method of removing small negatively charged organic molecules from a biological sample mixture, the method comprising:

providing a device comprising at least one process array that comprises a surface comprising quaternary ammonium ions partially coated with a negatively charged polyelectrolyte;

providing a biological sample mixture in the at least one process array; and

transferring the biological sample mixture within the at least one process array, wherein the biological sample mixture and the surface comprising quaternary ammonium ions partially coated with a negatively charged polyelectrolyte remain in contact for a sufficient time to remove at least a portion of the small negatively charged organic molecules from the biological sample mixture.

37. The method of claim 36 wherein the biological sample mixture comprises a nucleic acid amplification reaction mixture.

38. The method of claim 36 wherein the biological sample mixture and the surface comprising quaternary ammonium ions partially coated with a negatively charged polyelectrolyte are agitated while in contact.

39. A device for use in removing small negatively charged organic molecules from a biological sample mixture, the device comprising:

a plurality of process arrays, wherein each process array of the plurality of process arrays comprises:

a plurality of process chambers, each of the process chambers defining a volume for containing a biological sample mixture; and

at least one distribution channel connecting the plurality of process chambers;

a surface within at least one of the process arrays comprising an anion exchange material.

40. The device of claim 39 further comprising a plurality of valves, wherein at least one of the valves is located along the at least one distribution channel.

41. The device of claim 39 wherein the plurality of process arrays comprises a plurality of independent process arrays.

42. The device of claim 39 wherein the plurality of process arrays are arranged radially on the device.

43. The device of claim 39 wherein the surface comprising an anion exchange material comprises an anion exchange material partially coated with a negatively charged polymer.

44. The device of claim 43 wherein the negatively charged polymer is a polyelectrolyte.

45. The device of claim 43 wherein the anion exchange material, the negatively charged polymer, or both are pattern coated.

46. An analytical receptacle comprising one or more reservoirs and a surface with a cover film adhered to the surface and enclosing the one or more reservoirs; wherein the cover film comprises a backing and an adhesive disposed on at least one major surface of the backing and in contact with the receptacle surface; wherein at least a portion of the adhesive has an anion exchange material disposed thereon.

47. The analytical receptacle of claim 46 wherein the anion exchange material comprises quaternary ammonium ions.

48. The analytical receptacle of claim 46 wherein the anion exchange material is partially coated with a negatively charged polymer.

49. The analytical receptacle of claim 48 wherein the negatively charged polymer is a polyelectrolyte.

50. The analytical receptacle of claim 48 wherein the anion exchange material, the negatively charged polymer, or both are pattern coated.

51. An analytical receptacle comprising a plurality of reservoirs adapted for receipt of a biological sample mixture, wherein at least one reservoir comprises a surface comprising an anion exchange material partially coated with a negatively charged polymer disposed therein.

52. The analytical receptacle of claim 51 wherein the anion exchange material comprises quaternary ammonium ions and the negatively charged polymer is a negatively charged polyelectrolyte.